Effect of kiwiberry pre-storage treatments on the fruit quality during cold storage

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Abstract
The kiwiberry (Actinidia arguta (Siebold et Zucc.) Planch ex Miq.) has become a widely studied species in recent years due to its high resistance to low temperature and high content of health-promoting phytochemicals. The aim of this study was to verify the effect of pre-storage application of oxalic acid, salicylic acid and acetylsalicylic acid, calcium chloride (CaCl₂) and 1-methlycyclopropene (1-MCP) on quality changes of the kiwiberry cultivar ‘Weiki’ during cold storage. During six weeks of storage fruit firmness, titratable acidity and the total ascorbate content decreased from an average of 61 N, 1.5%, 857 mg kg⁻¹ to 2.5 N, 1%, 380 mg kg⁻¹, respectively. Contrary, the soluble solids and the total phenolics content increased from an average of 0.30%, 73.5 mg kg⁻¹ to 0.51%, 172 mg kg⁻¹, respectively. Application of 1-MCP clearly improved fruit firmness and inhibited the ascorbate loss. Compared to the control, the fruit firmness and total ascorbate content was higher by 50% and 13%, respectively. The acetylsalicylic acid treated fruit exhibited significant drop of phenolics as compared to the control. There was no significant difference in fruit total antioxidant activity determined immediately after harvest and at the end of storage (16.7 vs 17 mmol kg⁻¹, respectively). Fruit treated by salicylic acid, CaCl₂ and 1-MCP was characterized by a significantly higher titratable acidity compared to the control group. This study showed that the highest ability to maintain fruit quality during cold storage was exhibited by 1-MCP treated kiwiberry.

Key words: Actinidia arguta, antioxidant potential, ascorbate, mini kiwi, organic acids, phenolics, postharvest treatments.

Introduction
Actinidia arguta (Siebold et Zucc.) Planch ex Miq., also known as mini kiwi, kiwiberry, baby kiwi or hardy kiwifruit, names which all refer to small smooth fruit with edible green or red skin, is a promising fruit species, suitable for cultivation in temperate climate (Bieniek et al., 2016). Global commercial production of kiwiberry is continuously increasing (Latocha et al., 2015 b). The yield in 2015 was at least 500 tons (Latocha, Debersaques, 2016). In addition to being considered tasty with the ability to be eaten whole, kiwiberry was reported to contain large amounts of vitamin C, phenolics, carotenoids and some minerals (Nishiyama et al., 2004; 2005; Fisk et al., 2006; Bieniek, Dragańska, 2013; Latocha et al., 2015 a; b). In contrast to kiwifruit, kiwiberry peel can be considered as an additional source of those compounds (Kim et al., 2009; Latocha et al., 2015 b; Wojdylo et al., 2017). Furthermore, A. arguta plants show good winter hardiness as well as short growing and maturation period (Bieniek et al., 2016), hence, it can be grown in temperate climate zone. Apart from the above-described attributes, postharvest storage ability and fruit storage characteristics are of great importance, especially in the case of commercial production. Physiologically mature fruit of A. deliciosa cultivar ‘Hayward’ can be stored under refrigeration for 4–6 months, while the kiwiberry only 1–2 months (Fisk et al., 2006; Krupa et al., 2011; Latocha et al., 2014). Kiwiberry belongs to climacteric fruits in which ethylene speeds up the processes of fruit ripening and softening due to continued fruit respiration, which is accompanied by chlorophyll and starch degradation (Wang et al., 2015 a). Therefore, the application of ethylene action inhibitors is considered to be a promising method for the storage of climacteric fruit. Currently only fragmentary data exist on the effect of 1-MCP and other ethylene release inhibitors on the bioactive compound contents in kiwiberry (Wang et al., 2015 a).

Oxalic, acetylsalicylic and salicylic acids are compounds commonly present in plant tissue. It should be pointed out that as naturally occurring compounds they also satisfy consumer demands with respect to food preservation systems. It was reported that those
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Bioactive compounds and total antioxidant activity during cold storage.

Materials and methods

Material, harvest and storage conditions. Kiwiberry (Actinidia arguta (Siebold et Zucc.) Planch ex Miq.) cultivar ‘Weiki’ was harvested at commercial maturity stage (7–7.5% soluble solid content) (Latocha et al., 2014; Lim et al., 2016) from a commercial plot located in Bodzew, Mazowieckie State, Poland (51°47ʹ49.9″ N, 20°48ʹ44.0″ E). Fruit of a moderate and equal size, without any deformations or damage, was collected manually from a number of randomly selected plants on the 6th of September, 2014. Based on previous studies (Fattahi et al., 2010; Kazemi et al., 2011; Valero et al., 2011), the following pre-storage treatments were investigated: salicylic acid (SA), acetylsalicylic acid (ASA), oxalic acid (OX) and salicylic (SA) acids, calcium chloride (CaCl2) and widely used 1-methlycyclopropene (1-MCP) technique on the kiwiberry quality changes including the content of biologically active compounds as well as total antioxidant activity during cold storage.

Kiwiberry (Actinidia arguta (Siebold et Zucc.) Planch ex Miq.) cultivar ‘Weiki’ was harvested at commercial maturity stage (7–7.5% soluble solid content) (Latocha et al., 2014; Lim et al., 2016) from a commercial plot located in Bodzew, Mazowieckie State, Poland (51°47ʹ49.9″ N, 20°48ʹ44.0″ E). Fruit of a moderate and equal size, without any deformations or damage, was collected manually from a number of randomly selected plants on the 6th of September, 2014. Based on previous studies (Fattahi et al., 2010; Kazemi et al., 2011; Valero et al., 2011), the following pre-storage treatments were investigated: salicylic acid (SA), acetylsalicylic acid (ASA), oxalic acid (OX) – each of the compounds was used in the concentration of 2 mmol, next 2% calcium chloride (CaCl2) (w/v), 0.65 μL L⁻¹ 1-methlycyclopropene (1-MCP) and control (distilled water). Solutions were prepared in distilled water. In the control, SA, ASA and OX treatments, fruit was dipped in the solutions for 5 min. The fruit treated with 2% CaCl₂ was dipped for 2 min. Afterwards the fruit was dried at room temperature and packed into plastic punnets of 250 g capacity. Application of 1-MCP was made on fruit held in storage chambers (volume of one cubic meter). Fruit was gas-treated for 24 h at 1-MCP concentration of 665 ppm and temperature of 1°C. During this time fruit of other treatments was stored in contiguous chamber under the same temperature. Finally, the examined fruit was put into cold storage (1°C and 85% relative humidity) for 6 weeks. Storage chambers were equipped with automatic system “Oxystat 200” (David Bishop Ltd, Holland). The first fruit evaluation was made after 14 days of storage and next fruit was taken out for analysis every 7 days. In each term of measurement batch of fruit (20 fruit per replication, three replications per treatment) was transferred to the laboratory. Ten fruit from replication was used to analyse the basic physicochemical parameters. The other ten fruit was frozen in liquid nitrogen, pulverized to a fine powder and stored in −80°C until analysis of bioactive compounds and total antioxidant activity.

Fruit quality measurements. Fruit firmness. Instron 5542 (USA) with round probe 4.5 mm diameter at 4 mm s⁻¹ speed of compression was used to determine the fruit firmness. Assessments were performed on two opposite sides of each fruit. The results were expressed as maximum force to penetrate fruit to the depth of 8 mm in Newton (N). Fruit previously used for fruit firmness evaluation was homogenized and centrifuged at 5000 rpm and the supernatant was used for analyses of titratable acidity and soluble solid content. The soluble solid content was measured with the digital refractometer PR-32a (Atago, Japan). The results were expressed as % soluble solid content. The easy intelligent titrator TitroLine (SI Analytics GmbH, Germany) was used to measure titratable acidity. The kiwiberry juice (supernatant) was diluted with distilled water in the proportion 1:10 (v:v). The juice solution was then titrated with 0.1 M NaOH up to a pH value of 8.1 and expressed as citric acid equivalent (%).

Skin colour was measured on two opposite sides with a colorimeter CR-508i (Minolta, Japan), equipped with a 5 mm measuring head and observer 10° and illuminant D65. The meter was calibrated using the manufacturer’s standard white plate. Colour changes were quantified in the L*, a* and b* colour space. Hue angle (h°) was calculated from a* and b* values:

\[ h° = 180° \cdot \frac{b°}{a°} + 180° \text{, when a° < 0 and b° > 0,} \]

as well as chroma values:

\[ C = \sqrt{a^2 + b^2}. \]

Hue values refer to a colour wheel. Red, yellow, green and blue colours were at an angle of 0°, 60°, 120° and 240°, respectively. Chroma describes the vivdness or dullness of fruit colour and is also known as colour saturation.

Bioactive compounds and total antioxidant activity measurements. The total ascorbate content (tASC: the sum of L-AA and DHAA, reduced and oxidized forms, respectively) was measured using HPLC technique (Latocha et al., 2015 b). The powdered fruit tissue was suspended in 0.1 M HCl in a 1:30 (m/v) ratio. Next, samples were centrifuged at 14 000 rpm for 20 min at 4°C. The obtained extract was used for tASC determination. The tASC was measured after the complete oxidation of L-AA to DHAA with ascorbate oxidase. DHAA was then derivatized with o-phenylenediamine. The reaction product was detected fluorometrically at 450 nm by excitation at 250 nm. Separation was made on a SpherisorbR column (250 × 4.6 mm, 5μm, Waters) by applying a solution of 20% methanol containing 800 mM K2HPO4 (pH = 7.8), flow rate was 1 mL min⁻¹. The tASC was quantified using a calibration curve of L-ascorbic acid.

Total phenolics content was evaluated using the new method: Fast Blue BB (FBBB) procedure described by Medina (2011). Two-step extraction in ultrasonic bath was applied. The powdered fruit samples were extracted with 2.5 mL of 70% ethanol in a 1:10 (m/v) ratio in ultrasonic bath for 30 min and centrifuged at 20 000 rpm for 10 min at 4°C. The supernatant was decanted and transferred to glass tubes. The residue was extracted again with another 2.5 mL of 70% ethanol in the same conditions as described above. Next, the both extracts were combined, centrifuged (20 000 rpm, 10 min, 4°C), decanted to the Eppendorf tubes and stored in −20°C. Before measurement, the extracts were diluted with redistilled water in a 1:10 (v/v) ratio. FBBB reagent created coloured (blue) azo-complexes with phenolic compounds, which were detected spectrophotometrically
at 420 nm. The results were calculated using a calibration curve and expressed in gallic acid equivalents (GAE).

The FRAP-assay, referred to as total antioxidant activity, is a method based on the reduction of Fe3+-TPTZ (4,6-tri(pyridil-S-triazine) to a blue coloured Fe2+-TPTZ compound in the presence of antioxidants, which is revealed as a change of absorbance at 593 nm (Latocha et al., 2015 b). Samples were extracted with 5 mL of MilliQ water in a 1:10 (v/v) ratio and centrifuged at 14 000 rpm for 30 seconds. The obtained extract was filtered and the supernatant (diluted with MilliQ water in a 1:2 (v/v) ratio) was added to FRAP reagent containing 10 mM TPTZ in 40 mM HCl, 20 mM FeCl3, and 300 mM acetate buffer (pH = 3.6) in the ratio of 1:1:10. The FeSO4 compound was used to prepare the standard solutions.

Statistical analysis and presentation of the data. The obtained results were elaborated by two-way factorial analysis of variance (ANOVA) using software Statgraphics Plus 4.1 (USA). The significance of the differences between means of main effects (storage time and fruit pre-storage treatment) was evaluated using Tukey’s honestly significant difference (HSD) procedure, at 5% probability level. Therefore, the results were presented as mean figures with an indication of homological groups.

Results and discussion

Influence of fruit pre-storage treatment and time of storage on overall fruit quality. The fruit firmness loss is a physiological process that occurs during ripening, directly affecting postharvest life and the commercial value of different fruits. Short storability is a major problem for the kiwiberry industry, as the fruit firmness drops significantly during storage (Krupa et al., 2011). In this study, the fruit firmness was the only parameter for which the interaction between the main effects was significant (Table). After two weeks of storage fruit firmness of 1-MCP treated fruit was 43.7 N, while for the control group it was 11.7 N. Until the 4th week of storage, application of 1-MCP delayed the process of firmness loss considerably as compared to other treatments. The difference between 1-MCP and other treatments (based on their average fruit firmness values) reduced with time – from being 241% higher at 2nd week of storage to a 70% higher at 6th week of storage. Similarly, significant influence of 1-MCP on the fruit firmness of kiwiberry was recently described by Wang et al. (2015 a). In turn, in kiwifruit a delay of its softening during storage in 20°C was observed after SA application (Zhang et al., 2003). In other studies, the maintenance of Actinidia delicosa fruit firmness was achieved when fruit was immersed in CaCl2 solution for 1 and 5 minutes, respectively (Kazemi et al., 2011). Discriminants that influence the fruit taste in terms of quality and consumer acceptability are soluble solid content and titratable acidity.

During the first two weeks of storage, soluble solid content in fruit samples almost doubled (Fig. 1A). A smaller, but still significant, constant increase during the next two weeks was observed. Fruit from 4th to 5th week of storage showed the same soluble solid content, but near the end of storage period this quality parameter increased significantly again. There was no substantial influence of pre-storage treatments on soluble solid content (Fig. 1B). In this study, the soluble solid content and titratable acidity changes during storage were similar to those reported by Krupa et al. (2011). Summing up, this parameter decreased by approximately 54% during examined storage period (Fig. 1C). There was no significant difference between application of the ASA or OX compared to the control. In contrast, the SA, CaCl2, and 1-MCP treated fruit was characterized by a significantly higher values of titratable acidity compared to control group (Fig. 1D). Kazemi et al. (2011) reported that postharvest CaCl2 dips did not affect the titratable acidity in kiwifruit cultivar ‘Hayward’, while kiwifruit treated with SA solution achieved higher titratable acidity and lower soluble solid content than control fruit or those treated with CaCl2. In turn, a significant delaying effect of all analysed compounds (SA, ASA and OA) on the titratable acidity and soluble solid content in sweet cherry fruit was recently described (Valero et al., 2011). Hue and chroma values considerably decreased during storage, indicating a great loss of colour intensity (Fig. 1E, 1G). Compared to the results obtained after harvest a significant decrease of chroma occurred after two weeks of fruit storage (Fig. 1G), while hue angle after 4 weeks of storage (Fig. 1E). Hue values declined more gradually than chroma, which dropped more rapidly. Hue angle remained on the same level regardless of pre-storage treatment (Fig. 1F) while the CaCl2, treated fruit had a significantly higher chroma values as compared to the control, OX and ASA treatments (Fig. 1H). Although the decrease of hue and chroma values was clearly visible during fruit storage, there were no significant effects of examined treatments on the fruit colour changes in this study. Only the use of CaCl2 favoured a higher green colour saturation of the kiwiberry peel. Whereas the positive influence of 1-MCP treatment on pulp colour of kiwifruit was described by Boquete et al. (2004).

### Table. Effect of treatment and time of storage on Actinidia arguta fruit firmness

<table>
<thead>
<tr>
<th>Pre-storage treatment (B)</th>
<th>0</th>
<th>2nd</th>
<th>3rd</th>
<th>4th</th>
<th>5th</th>
<th>6th</th>
<th>Average (B)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>61.0 ± 2.0</td>
<td>11.7 ± 0.9</td>
<td>5.2 ± 0.2</td>
<td>3.2 ± 0.2</td>
<td>2.9 ± 0.3</td>
<td>2.1 ± 0.2</td>
<td>14.4 a</td>
</tr>
<tr>
<td>ASA</td>
<td>61.0 ± 2.0</td>
<td>13.9 ± 1.1</td>
<td>5.8 ± 0.3</td>
<td>3.4 ± 0.1</td>
<td>2.9 ± 0.5</td>
<td>2.2 ± 0.4</td>
<td>14.9 a</td>
</tr>
<tr>
<td>SA</td>
<td>61.0 ± 2.0</td>
<td>13.0 ± 0.4</td>
<td>5.2 ± 0.4</td>
<td>4.1 ± 0.2</td>
<td>3.0 ± 0.1</td>
<td>2.2 ± 0.2</td>
<td>14.7 a</td>
</tr>
<tr>
<td>OX</td>
<td>61.0 ± 2.0</td>
<td>13.1 ± 0.7</td>
<td>5.3 ± 0.2</td>
<td>3.2 ± 0.4</td>
<td>2.8 ± 0.1</td>
<td>2.4 ± 0.2</td>
<td>14.6 a</td>
</tr>
<tr>
<td>CaCl2</td>
<td>61.0 ± 2.0</td>
<td>12.2 ± 0.9</td>
<td>5.3 ± 0.3</td>
<td>3.6 ± 0.1</td>
<td>3.1 ± 0.5</td>
<td>2.2 ± 0.1</td>
<td>14.6 a</td>
</tr>
<tr>
<td>1-MCP</td>
<td>61.0 ± 2.0</td>
<td>43.7 ± 2.5</td>
<td>10.4 ± 0.4</td>
<td>6.4 ± 0.1</td>
<td>4.7 ± 0.1</td>
<td>3.8 ± 0.6</td>
<td>21.7 b</td>
</tr>
<tr>
<td>Average (A)</td>
<td>61.0 e</td>
<td>17.9 d</td>
<td>6.2 c</td>
<td>4.0 b</td>
<td>3.2 ab</td>
<td>2.5 a</td>
<td></td>
</tr>
</tbody>
</table>


The values are means of three replications, ±SD. Values in the row (means for consecutive storage times) or column (means for pre-storage fruit treatment) marked with different letter differ significantly at P ≤ 0.05 (Tukey HSD test).
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Notes. SA – salicylic acid, ASA – acetylsalicylic acid, OX – oxalic acid, CaCl₂ – calcium chloride, 1-MCP – 1-methylocyclopropene. The values are means obtained for a given level of main effects (weeks of storage or fruit pre-storage treatment), ±SD. Values marked with different letters differ significantly at $P \leq 0.05$ (Tukey HSD test); ns – not significant.

Figure 1. Effect of storage time (A, C, E, G) and treatment (B, D, F, H) on soluble solids content, titratable acidity and skin colour (hue angle and chroma) in *Actinidia arguta* fruit

*The tASC and total phenolics content contents and total antioxidant activity (FRAP-assay): the effect of storage time and fruit pre-storage treatment.* Actinidia genus is described as a rich source of biologically active compounds (Nishiyama et al., 2004; 2005; Latocha et al., 2015 b). Maintaining or enhancing fruit internal quality is the second, after storability potential, important issue in relation to post-harvest fruit management (Fisk et al., 2006; Xia et al., 2016). The extent of fluctuations in the content of phytochemicals during storage depends not only on the storage conditions and pre-storage treatment, but also chemical nature of the compounds and fruit type.
(Łata, 2008; Rapisarda et al., 2008). A general agreement among researchers exists, that fruit phenolic metabolism remains relatively low during long-term storage whereas ascorbic acid is more sensitive to degradation due to its high oxidative capability during food storage or other postharvest treatments (Kevers et al., 2007; Łata, 2008). However, looking through individual studies, the pattern of these compounds changes during storage and shelf life might be sometimes variable and contradictory (Kalt, 2005; Amodio et al., 2007; Jhalegar et al., 2011). The kiwiberry tASC after harvest was measured as 857 mg kg\(^{-1}\) fresh weight and its concentration decreased significantly during storage regardless of treatment type (Fig. 2A). At the end of experiment the tASC was from 1.6 to 2.6 times lower for 1-MCP and control fruit as compared to harvest, respectively (data not shown). Decrease in vitamin C content in \emph{A. arguta} fruit during 42 days of cold storage was also reported by Krupa et al. (2011). In contrast, Kazemi et al. (2011) described that ascorbic acid level in \emph{A. delicosa} fruit increased during two months of storage at 1°C, and the SA and CaCl\(_2\), treated fruit showed significantly higher concentration of vitamin C compared to the control. In our study, compared to the control, tASC losses were significantly reduced after 1-MCP treatment (Fig. 2B). However, according to Vilaplana et al. (2006) ascorbate pathway is not ethylene dependent and therefore 1-MCP treatment should not affect considerably ascorbate status. Besides 1-MCP, some positive effect on tASC occurred after application of OX and CaCl\(_2\) compounds. Other than having high vitamin C content, kiwiberry is also considered as a rich source of phenolic compounds, richer than that of commonly eaten kiwifruit or apple (Kim et al., 2009). In general, total phenolics content successively increased during storage (Fig. 2C). After the first three weeks of storage the total phenolics content was similar to that obtained after harvest. Especially high increase of total phenolics content was noted after four weeks of cold storage. At the 5\(^{th}\) week, the rate of total

**Notes.** SA – salicylic acid, ASA – acetylsalicylic acid, OX – oxalic acid, CaCl\(_2\) – calcium chloride, 1-MCP – 1-methylcyclopropene. The values are means obtained for a given level of main effects (weeks of storage or fruit pre-storage treatment), \(\pm SD\). Values marked with different letters differ significantly at \(P \leq 0.05\) (Tukey HSD test); ns – not significant.

**Figure 2.** Effect of storage time (A, C, E) and treatment (B, D, F) on the total ascorbate content (tASC), total phenolics content and total antioxidant activity (FRAP-assay) in fresh weight of \emph{Actinidia arguta} fruit.
phenolics content increase however started to reduce. The total phenolics content after the 4 and 5 weeks of storage was approximately 46% higher compared to the content at harvest time. The above-described general picture of total phenolics content changes was nearly the same for all treatments up to the 4th week. After that time, fluctuations in total phenolics content were more treatment dependent. It was noted that the type of the fruit pre-storage treatment influenced total phenolics content to a lesser degree than storage time (Fig. 2D). There is a possibility that some biochemical changes in fruit could occur in part due to transpiration of moisture during storage; however, no more than 2.3% of fruit mass was lost due to transpiration after 6 weeks of storage (data not shown). The increase of total phenolics content up to 35 days of storage was also observed by Amodio et al. (2007) in kiwifruit. Zorić et al. (2017) noted that anthocyanins were more susceptible to degradation compared to other phenolic compounds in stored fruit. Recently, twenty one different phenolic compounds were detected and identified in A. arguta cultivar ‘Weiki’ fruit by Wojdyło et al. (2017), including flavonols, phenolic acids and anthocyanins. The total phenolics content was reported as 3691 mg 100 g⁻¹ dry weight. Authors stated that in A. arguta fruit anthocyanins present a very marginal percent of all phenolic compounds (Wojdylo et al., 2017). Valero et al. (2011) suggested that some increase in phenolics and anthocyanins concentrations during storage period is due to the postharvest ripening process. An increase in anthocyanins, flavonones and hydroxycinnamic acids and a slight decrease in vitamin C in the blood oranges during storage was noted (Rapisarda et al., 2008). In turn, cold storage negatively affected flavanone concentration. Other study (Hallborth et al., 2006) demonstrated that the selected enzymes from the flavonoid pathway that affect flavonoid biosynthesis show two distinct activity peaks during fruit ripening at early and late developmental stages. The first activity peak corresponds to the formation of flavanols, while the second peak is clearly related to anthocyanin and flavonol accumulation.

However, contrary to our study, a gradual decrease of total phenolics content in kiwifruit during 5 weeks of cold (1°C) storage after 1-MCP treatment was reported by Lim et al. (2016). Krupa et al. (2011) also noted decreasing total phenolics content levels during cold storage in A. arguta fruit but with no pre-storage treatment. Liu et al. (2015) reported that 1-MCP could delay or reduce the total phenolics content accumulation in peaches, which is in agreement with studies on avocados (Zhang et al., 2013). In this study, as compared to the control and other treatments, no particular effect of 1-MCP on total phenolics content was noted. The highest total phenolics content was noted in fruit after pre-storage OX application. Moreover, fruit treated by ASA was characterized be a significantly lower total phenolics content as compared to the control. Results recently obtained by Zhu et al. (2016) were promising in relation to OX application. Authors stated that the OX treatment increased quality and induced disease resistance in kiwifruit. Summing up, it is difficult to outline a clear pattern of phenolic changes, phenol content may either increase or decrease depending on the species, the storage conditions or fruit postharvest (pre-storage) treatment.

Ascorbate and phenolics as well as other antioxidants contribute to the overall fruit total antioxidant activity. There were no significant differences between total antioxidant activity determined immediately after harvest and at the end of fruit storage (Fig. 2E). However, it should be added that between these two extreme point of measurements, in sequence, increases (2nd, 4th and 6th week) and decreases (3rd and 5th week) of the total antioxidant activity, compared to harvest time, were noted. The highest drop of fruit total antioxidant activity was measured between 4th and 5th week. Compared to the second week, in which FRAP value was the highest during kiwiberry storage, the total antioxidant activity after 5 weeks of storage decreased by 34%. The examined treatments did not differ significantly in total antioxidant activity (Fig. 2F). Shivashankara et al. (2004) suggested that an increase in antioxidant capacity during cold storage may be possible only in fruit in which the contribution of total phenolics to total antioxidant activity is greater than that of the ascorbic acid. Tavarini et al. (2008) reported the correlation studies on antioxidants and phytochemical constituents responsible for antioxidant capacity in kiwifruits. Authors suggested that in kiwifruits vitamin C contributed to antioxidant capacity much more than other antioxidant constituents (phenols or carotenoids). However, in Actinidia genus a decrease (Tavarini et al., 2008; Krupa et al., 2011) or an increase (Amodio et al., 2007; Bucheri et al., 2015) of total antioxidant activity during cold storage was noted. The above mentioned inconsistency might be caused by various content and behaviour of individual bioactive compounds during storage and some differences in storage conditions or the way of measurement of total antioxidant activity (Lata, 2008). In our study, the total antioxidant activity could have remained almost unchanged probably because of a simultaneous rise of the total phenolics content and significant decrease of the tASC.

Conclusions

1. According to our study, 1-methycyclopropene (1-MCP) postharvest treatment had significant effect on kiwiberry (Actinidia arguta) ripening, manifesting itself by delaying the decline in fruit firmness and retarding the increase of soluble solids content.
2. 1-MCP as well as calcium chloride (CaCl₂) and salicylic acid (SA) significantly delayed the decrease in the titratable acidity compared to the control.
3. Regardless of the postharvest treatment method used, total phenolics content considerably increased but total ascorbate content (tASC) significantly decreased during cold storage.
4. The pre-storage treatment with acetylsalicylic acid (ASA) significantly reduced the total phenolics content in kiwiberry fruit. In turn, oxalic acid (OX) application apparently, but not significantly, increased the phenolics content.
5. Application of 1-MPC, followed by OX and CaCl₂, maintained the kiwiberry vitamin C content at the highest level during cold storage.
6. The examined treatments did not have significant influence on the total antioxidant activity of kiwiberry.
7. The possible solutions to improve quality characteristics during storage may involve combining treatment methods, changing treatment times or changing concentration of applied compounds.

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Smailialapės aktinidijos vaisių apdorojimo prieš sandėliavimą įtaka vaisių kokybei juos laikant šaltai

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Santrauka

Pastaraisiais metais smailialapė aktinidija (Actinidia arguta (Siebold et Zucc.) Planch ex Miq) tapo plačiai tyrimėjama rūšimi dėl jos aptarumo žemoms temperatūroms ir didelio kiekio sveikatai naudingu biologiškai aktyvių medžiagų. Tyrimo tikslas – nustatyti oksalo, salicilo ir acetilsalicilo rūgščių, kalcio chlorido (CaCl2) ir 1-metilciklopropeno (1-MCP) įtaką veislės ‘Weiki’ aktinidijų vaisių, apdorotų prieš sandėliavimą, kokybės pokyčiams juos laikant šaltai. Per 6 sandėliavimo savaites vaisių kietumas, titruojamasis rūgštumas ir suminis askorbatų kiekis sumažėjo vidutiniškai nuo 61 N, 1,5 %, 857 mg kg-1 iki 2,5 N, 1 %, 380 mg kg-1, o tirpiųjų kietųjų medžiagų ir suminis fenolių kiekis padidėjo vidutiniškai atitinkamai nuo 0,30 %, 73,5 mg kg-1 iki 0,51 %, 172 mg kg-1. Apdorojimas 1-MCP pagerino vaisių kietumą ir sumažino askorbatų netekimą. Palyginus su kontroliniu variantu, vaisių kietumas ir suminis askorbatų kiekis buvo atitinkamai 50 ir 13 % didesni. Vaisiuose, apdorotose acetilsalicilo rūgštimi, smarkiai sumažėjo fenolių, palyginus su kontroliniu variantu. Suminis antioksidacinis aktyvumas esmingai nesiskyrė tarp vaisių skynimo datų, nepriklausomai nuo apdorojimo (atitinkamai 16,7 ir 17 mmol kg-1 skynimo metu ir sandėliavimo pabaigoje). Vaisiai, apdoroti salicilo rūgštimi, CaCl2 ir 1-MCP, pasižymėjo esmingai didesniu titruojamuoju rūgštumu, palyginus su kontroliniu variantu. Tyrimo duomenys parodė, kad laikant šaltai geriausios kokybė išliko aktinidijos vaisiai, apdoroti 1-MCP.

Reikšminiai žodžiai: Actinidia arguta, antioksidacinis potencialis, askorbatas, fenoliai, mažieji kiviai, organinės rūgščios, nuskintų vaisių apdorojimas.